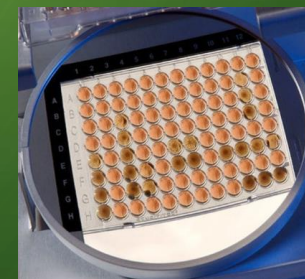


# *OFFICIAL SURVEILLANCE ON AMR BY CLASSICAL AND MOLECULAR METHODS IN LUXEMBOURG*

M. MEO<sup>1</sup>, D. CLAUDE<sup>2</sup>, M. PERRIN<sup>1</sup>, S. LOSCH<sup>2</sup>

<sup>1</sup>LABORATOIRE NATIONAL DE SANTÉ, 1 RUE LOUIS RECH, 3555 DUDELANGE

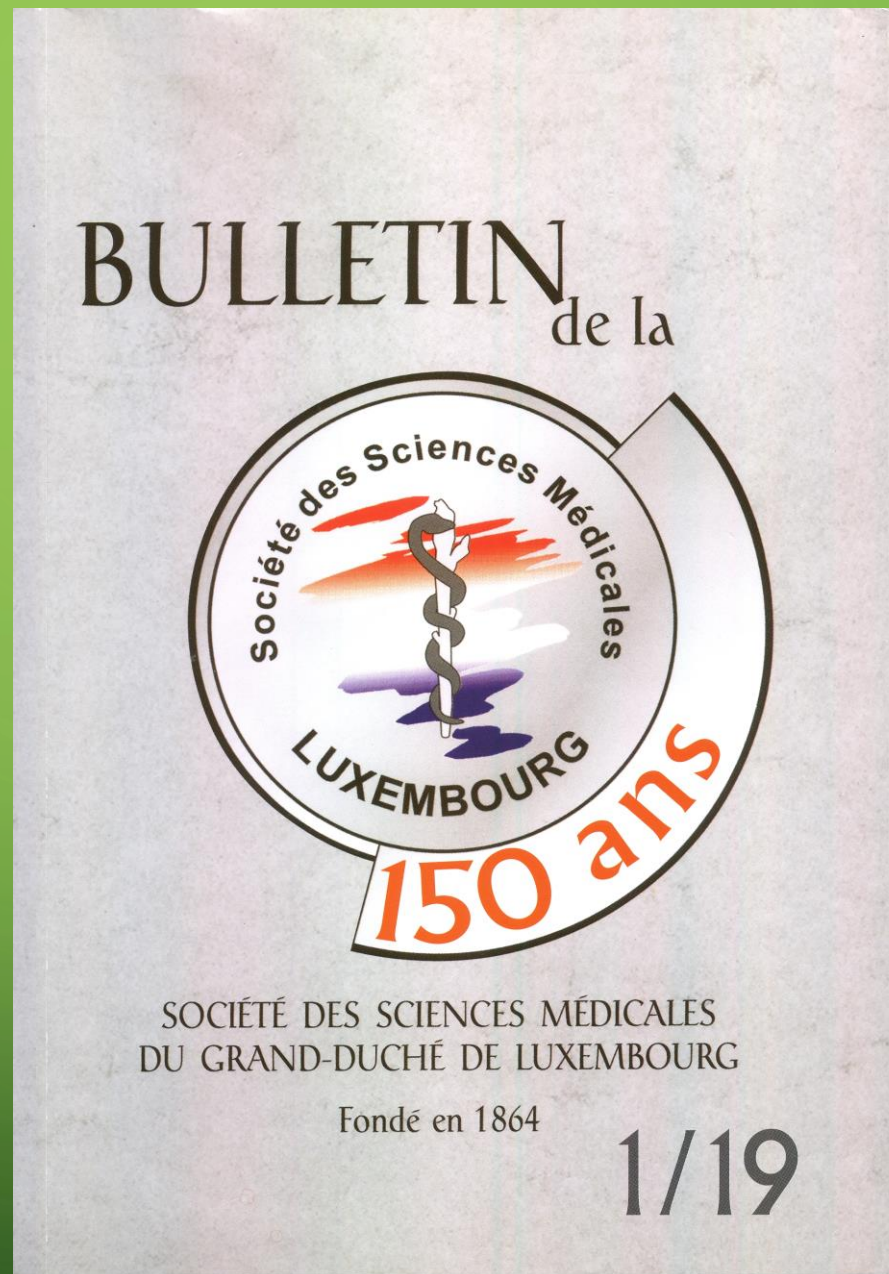
<sup>2</sup>LABORATOIRE DE MÉDECINE VÉTÉRINAIRE DE L'ÉTAT, 1 RUE LOUIS RECH, 3555 DUDELANGE



Data published:

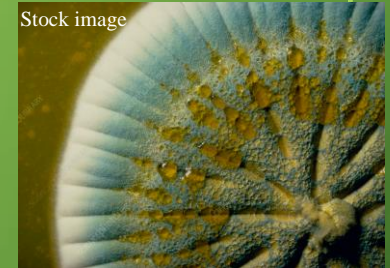
Meo M., Claude D., Perrin M.,  
Losch S.

2019, Bulletin de la Société des  
Sciences médicales 1/19, p.17-38



# OVERVIEW

- ❖ Introduction
- ❖ Material and Methods
- ❖ Target bacteria
- ❖ Results
- ❖ Discussion
- ❖ Conclusions



# MATERIAL & METHODS : SAMPLES



## Food:

- Any pure meat from beef, pork & poultry
- Sampling by official vets
- Bias by EU program (Decision 2013/652/EU)

## Animals:

- Feces, organs, milk
- Decision 2013/652/EU
- Clinical or necropsy samples
- Sampling by practitioners or official vets



## Clinical human samples:

- European surveillance of gastroenteric pathogens
- Collection by clinical labs → LNS
- ESBL transfer voluntarily

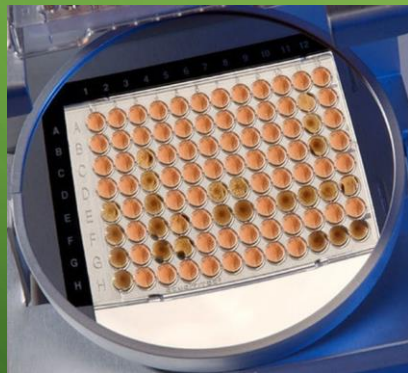


# MATERIAL & METHODS



Culture → isolation → antibiogram

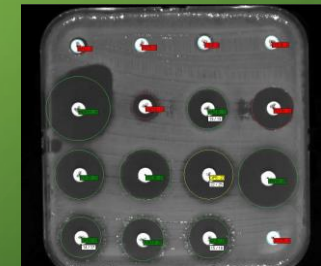
id. LMVE + ready strains → abg



Sensititre ®



**Interpretation:** CLSI, SFM, EUCAST



EUCAST

## MATERIAL & METHODS

- Sample collection by clinical labs transfer to LNS for ESBL typing
- By multiplex PCR from Dallenne *et al.* and comparison of PCR product migration on agarose gel.
- wgs

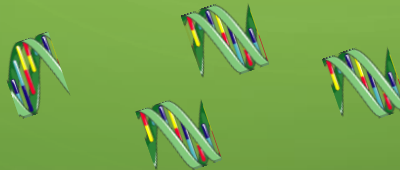


Culture



DNA  
extraction

PCR : ESBL gene research  
and amplification



## Molecular methods for ESBL

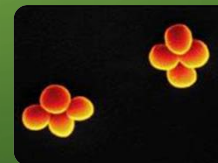
# TARGET BACTERIA (NAP)



Isolates from stool (LNS)  
food / faeces or organs (LMVE)



- *Campylobacter jejuni & coli*  
(→ n°1 food borne disease EU and L: bloody diarrhea, complications)
- *Salmonella*  
(→ n°2 food borne disease EU and L: diarrhea..., more or less severe illness)
- *Escherichia coli*  
(friend or enemy; diverse diseases if pathogenic)
- (*Staphylococcus aureus*)  
(→ food poisoning, skin or systemic diseases)

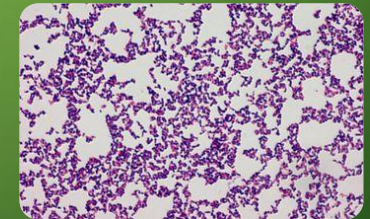


→ *comparison: human vs. non human of these zoonotic agents*  
**data 2016 – 2018**

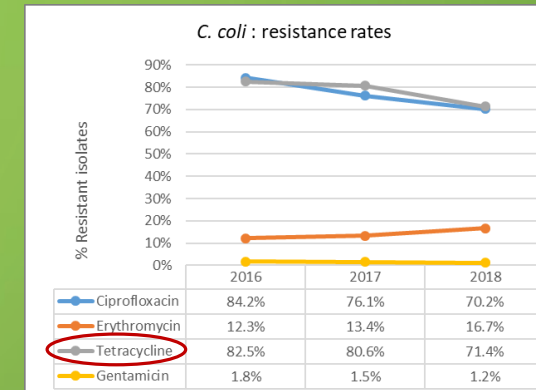
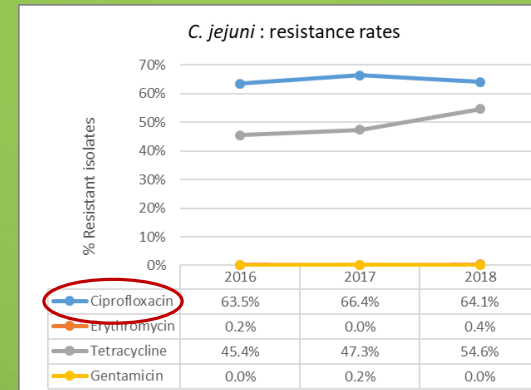
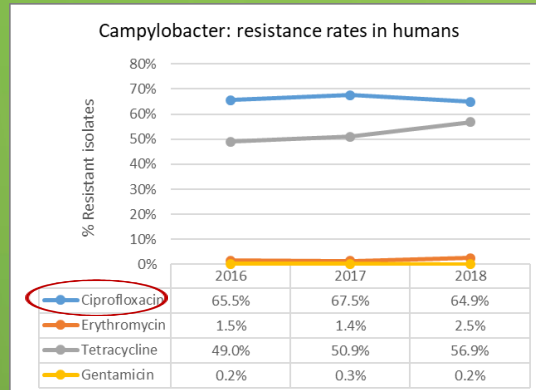
# TARGET BACTERIA “BULLETIN”



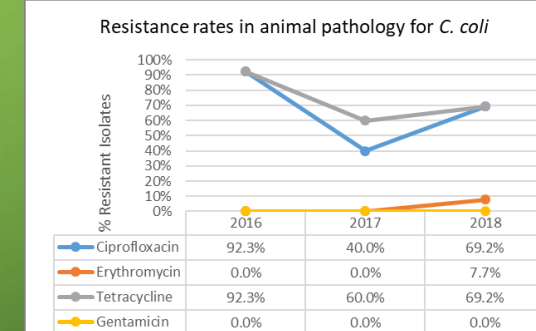
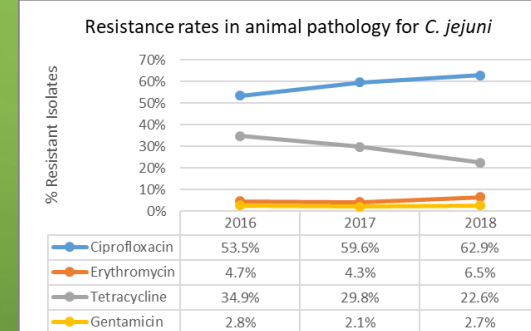
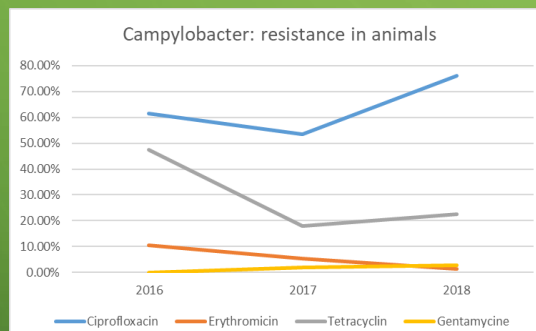
- *Pasteurella multocida*: Zoonotic agent ( → pulmonary diseases)  
isolated from milk or lungs
- *Trueperella pyogenes*: → suppurative mastitis, pneumonia in  
ruminants, pigs, horses, poultry and other species,
- *Campylobacter hyointestinalis* → proliferative enteritis  
Isolated from bovine faeces
- *Staphylococcus aureus*: **milk vs food**  
→ skin, udder or systemic diseases, food poisoning



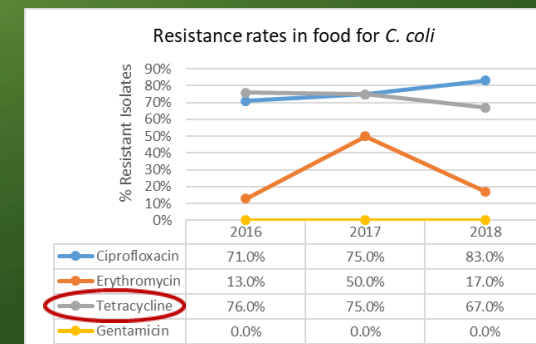
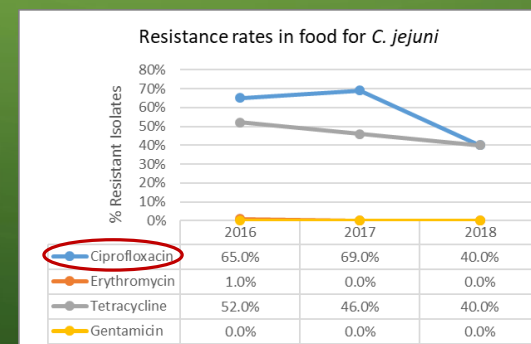
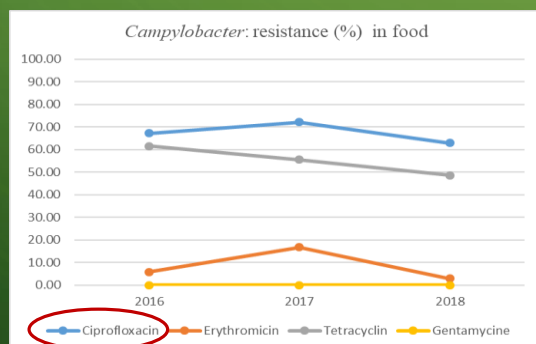
# RESULTS: *CAMPYLOBACTER*



1700 strains



188 strains



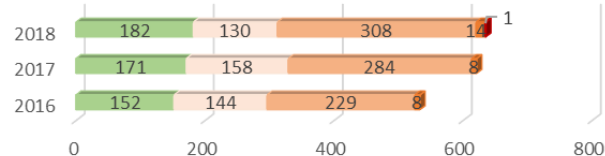
162 strains

## RESULTS : *CAMPYLOBACTER*

# MULTIDRUGRESISTANCE

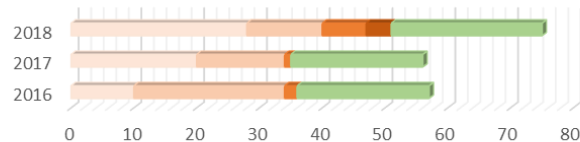


Multi Drug Resistant *Campylobacter*



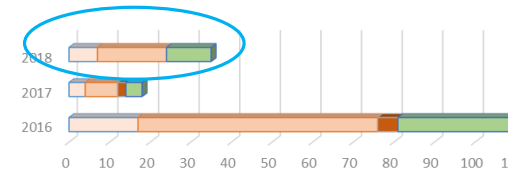
|       | 2016 | 2017 | 2018 |
|-------|------|------|------|
| S     | 152  | 171  | 182  |
| 1 AMR | 144  | 158  | 130  |
| 2 AMR | 229  | 284  | 308  |
| 3 AMR | 8    | 8    | 14   |
| 4 AMR |      |      | 1    |

*Campylobacter*: multiresistant animal pathology strains



|       | 2016 | 2017 | 2018 |
|-------|------|------|------|
| 1 AMR | 10   | 20   | 28   |
| 2 AMR | 24   | 14   | 12   |
| 3 AMR | 2    | 1    | 7    |
| 4 AMR | 0    | 0    | 4    |
| S     | 21   | 21   | 24   |

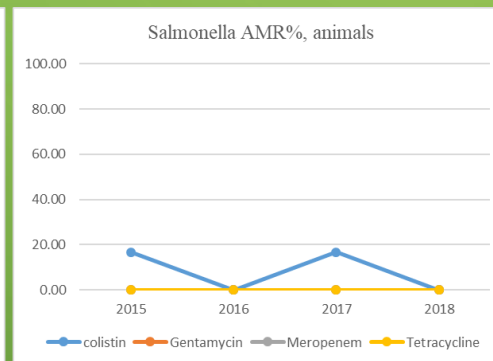
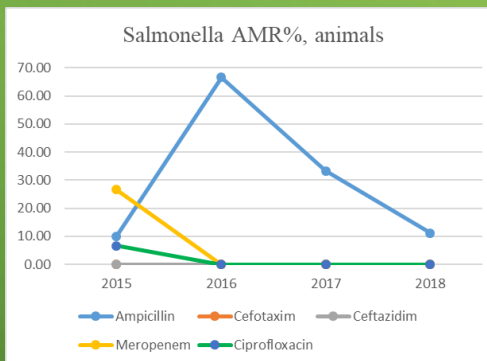
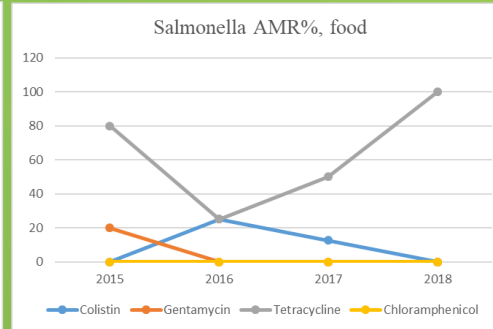
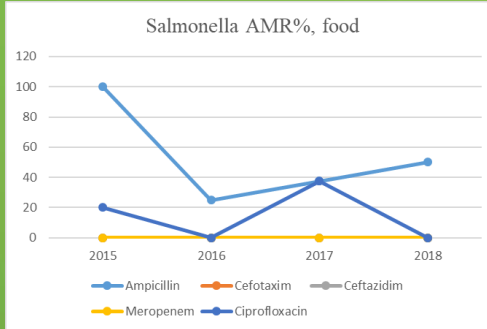
*Campylobacter*: multiresistant food strains



|       | 2016 | 2017 | 2018 |
|-------|------|------|------|
| 1 AMR | 17   | 4    | 7    |
| 2 AMR | 59   | 8    | 17   |
| 3 AMR | 5    | 2    | 0    |
| 4 AMR | 0    | 0    | 0    |
| S     | 28   | 4    | 11   |

6 ab tested

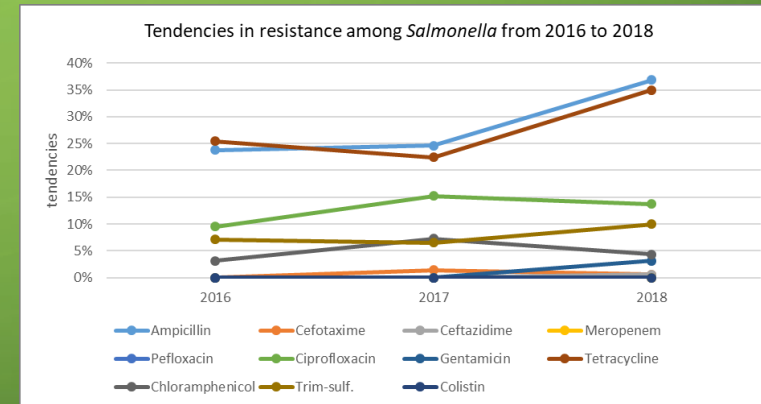
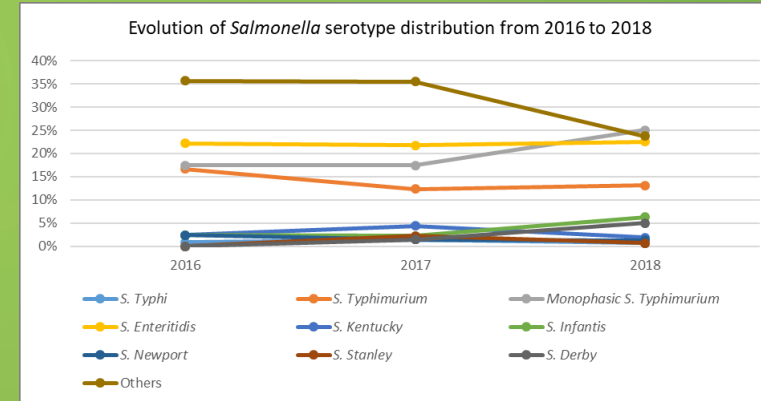
# RESULTS: *SALMONELLA*



19 food + 53 animal strains tested 2015-2018  
14 food + 23 animal strains tested 2016-2018

Colistin resistant *S.* in food:  
Enteritidis (2016), Typhimurium (2017)

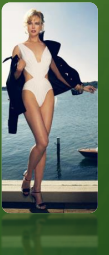
Colistin resistant *S.* in animals:  
Enteritidis, Give, Livingston (2015)  
Dublin (2019)



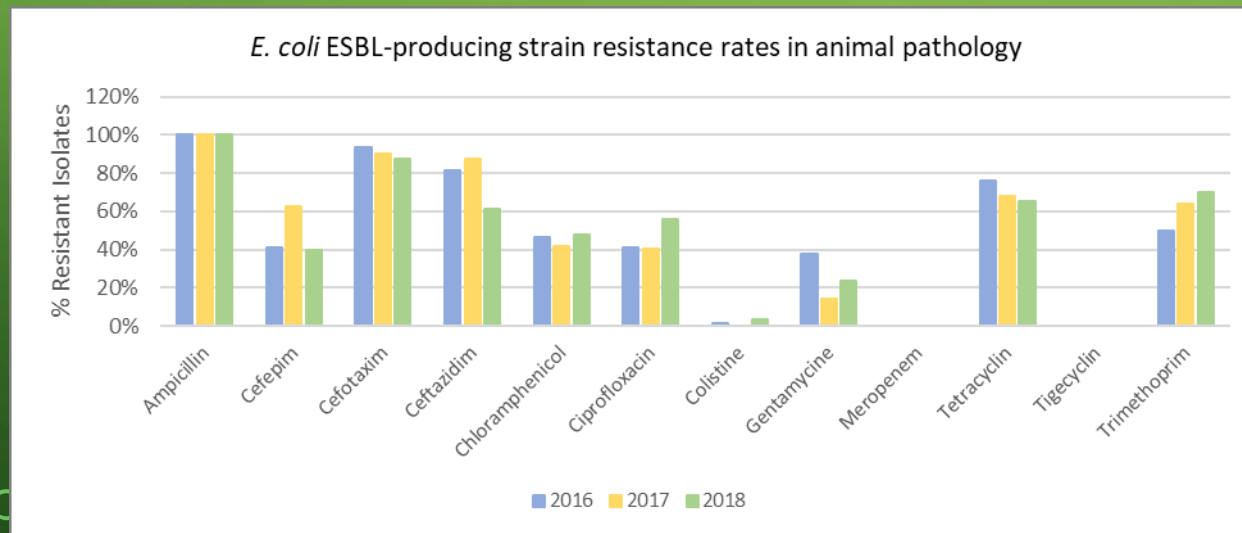
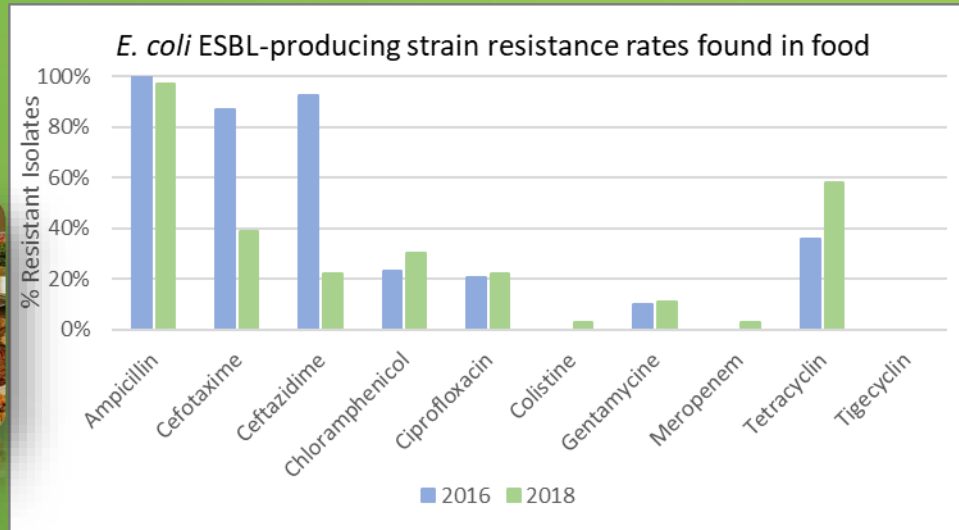
*S. enteritidis*, *S. Typhimurium* (bi-mono ph.) !!!

*S. Kentucky*, *Infantis*, *Derby*, *Newport*, *Stanley*  
Every year isolated

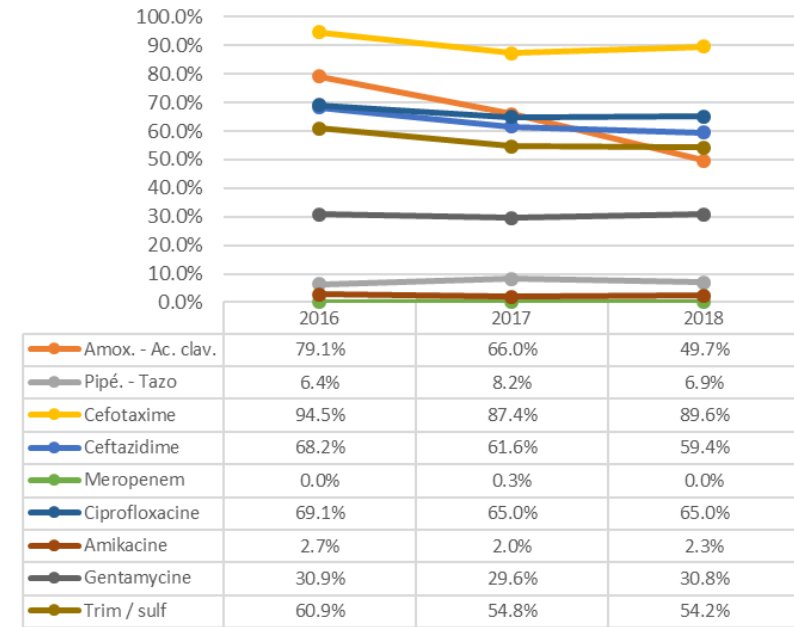
424 strains



# RESULTS: *E. COLI* ESBL



*E. coli* ESBL : resistance rates in humans



CTX-M 1,9; SHV, TEM, OXA-1 like  
most frequent genes  
Associations → MDR

2016-2018

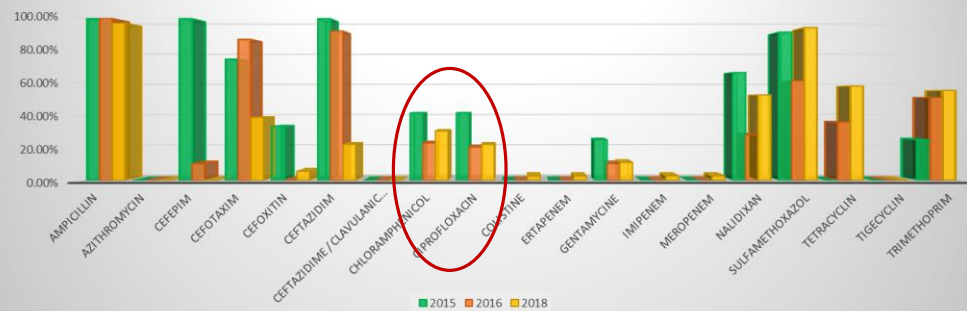


# RESULTS: *E. COLI* ESBL

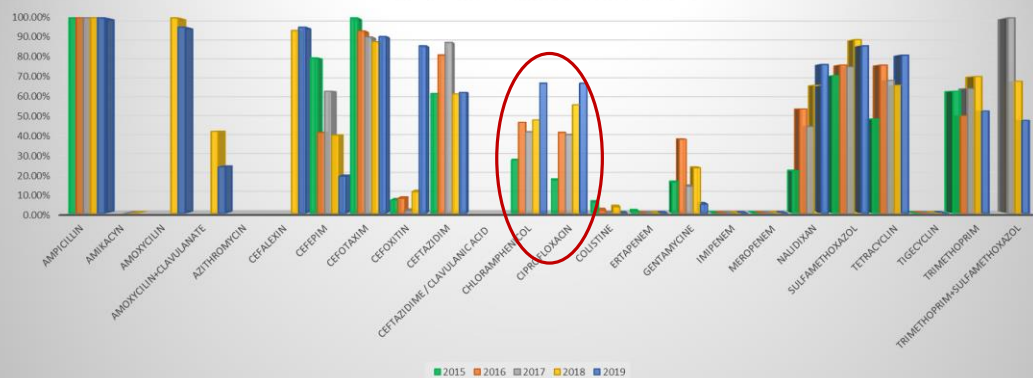
*E.coli* ESBL isolates tested

|         | 2015 | 2016 | 2017 | 2018 | 2019 |
|---------|------|------|------|------|------|
| food    | 12   | 39   | 3    | 36   | 3    |
| animals | 88   | 58   | 72   | 90   | 28   |

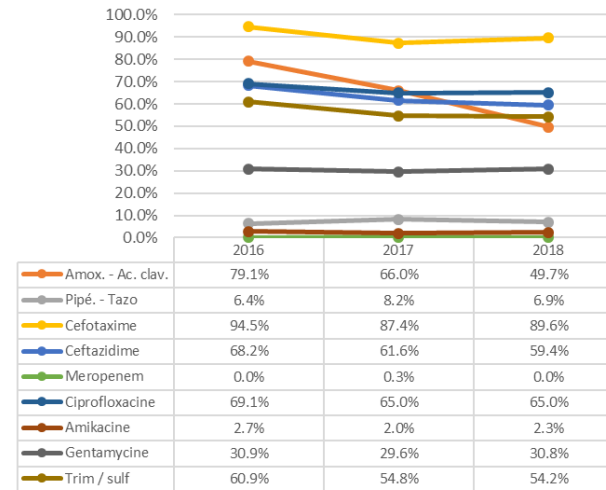
*E. coli* ESBL: resistance rate in food



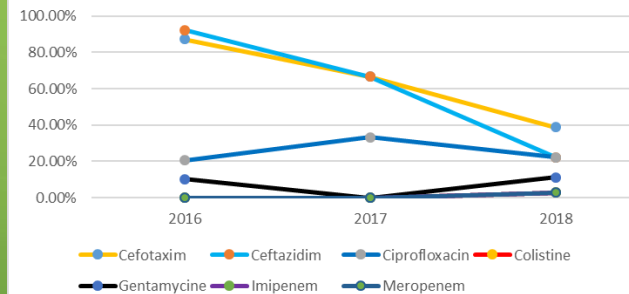
*E. coli* ESBL: resistance rate in animals



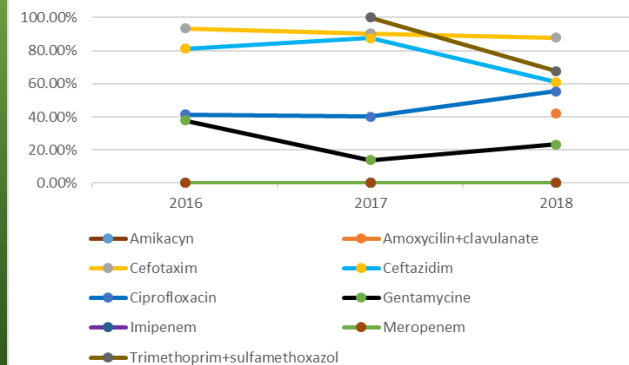
*E. coli* ESBL : resistance rates in humans



*E. coli* ESBL resistance rates in food



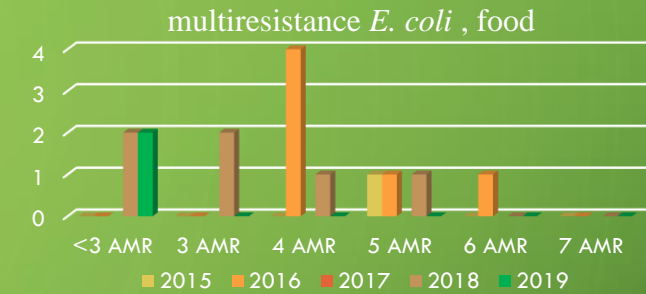
*E. coli* ESBL: resistance rates in animals



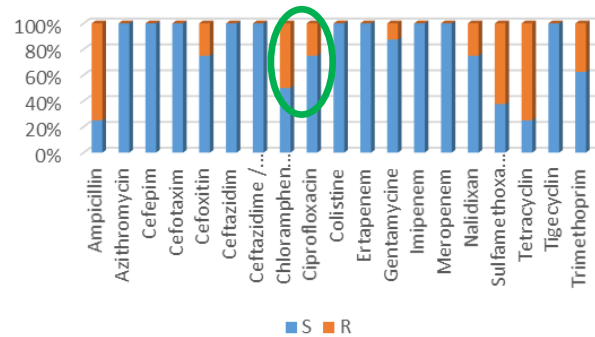
# RESULTS: *E. COLI*



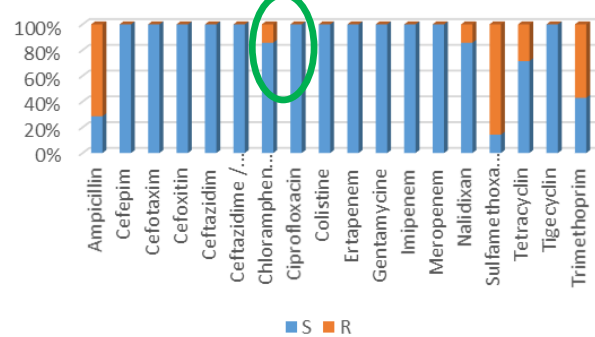
## MDR



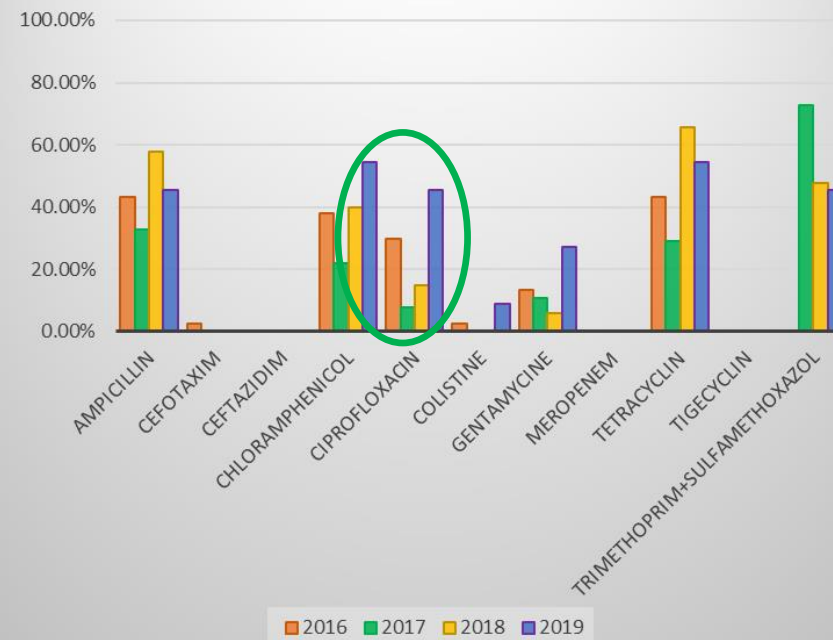
*E. coli* 2016, food



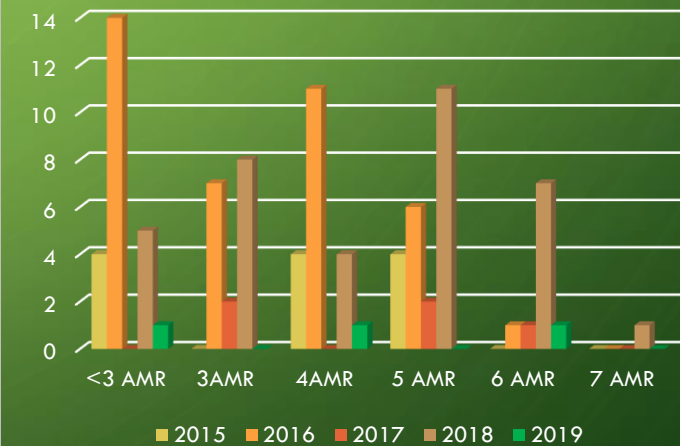
*E. coli* 2018, food



*E. coli*, animals

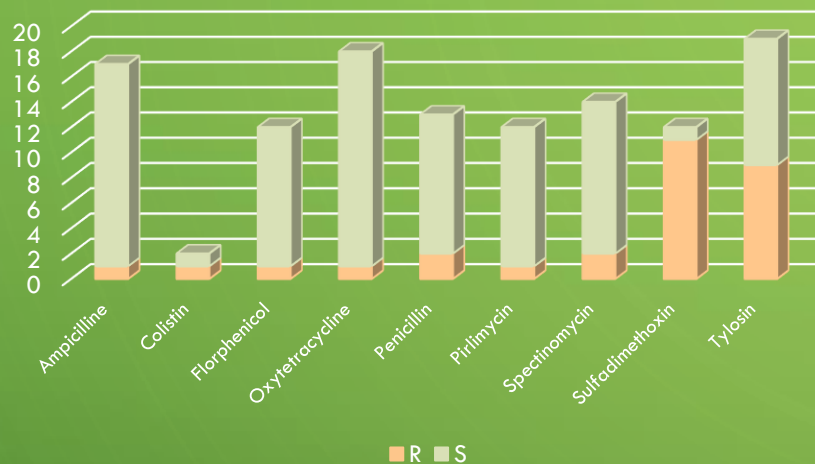


multiresistance *E. coli* ESBL, food

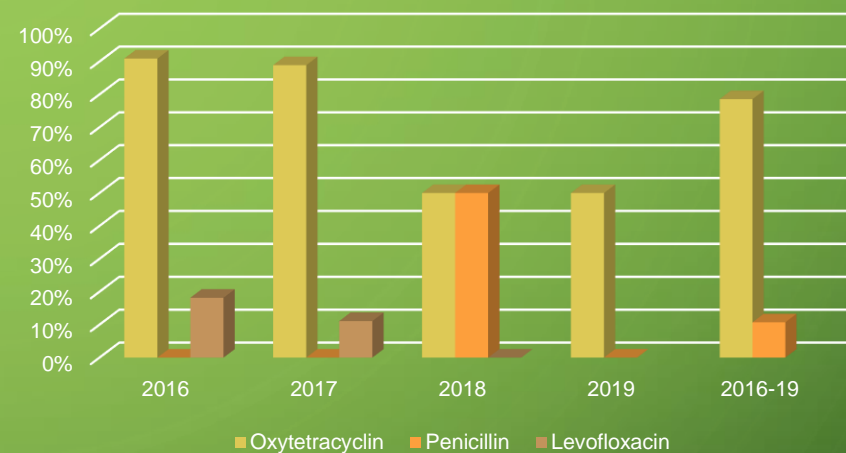


# SPECIAL VETERINARY STRAINS:

AMR for *Pasteurella multocida* from 2016 to 2019

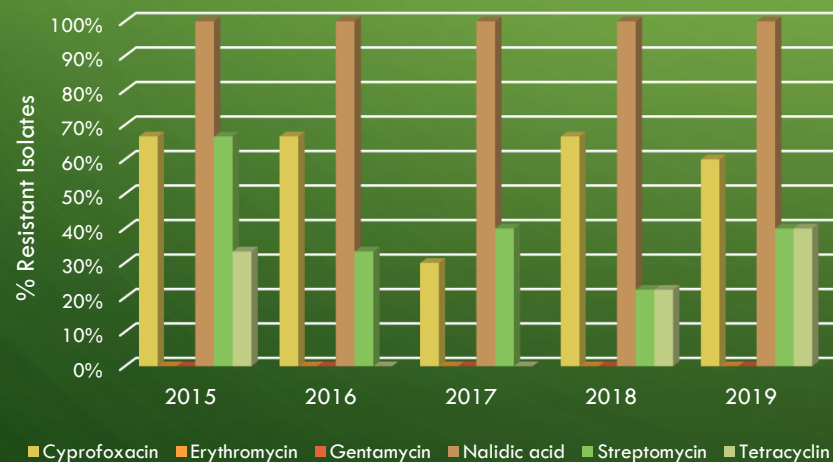


AMR for *Trueperella pyogenes* from 2016 to 2019

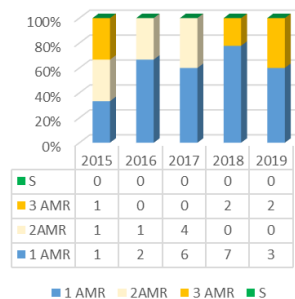


2/39 MDR

AMR for *C. hyointestinalis*



*Campylobacter hyointestinalis*  
MDR

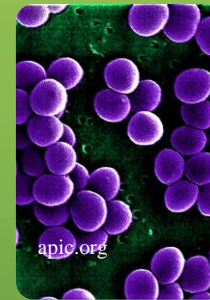
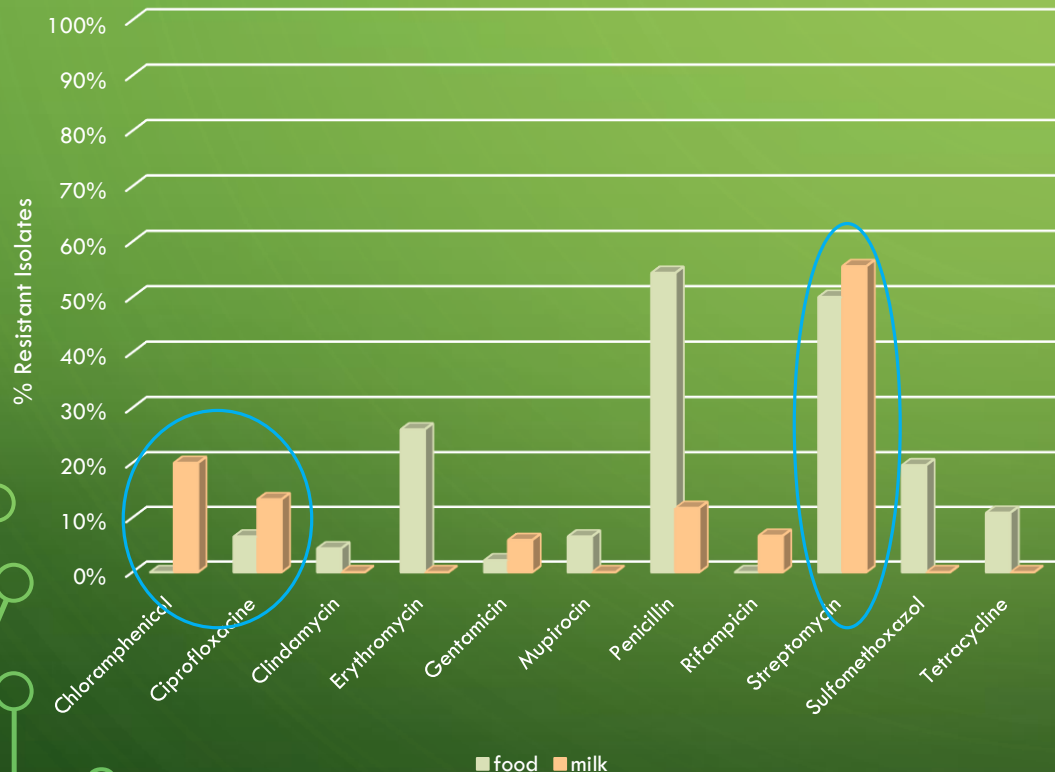


# SPECIAL VETERINARY STRAINS: *STAPHYLOCOCCUS AUREUS*

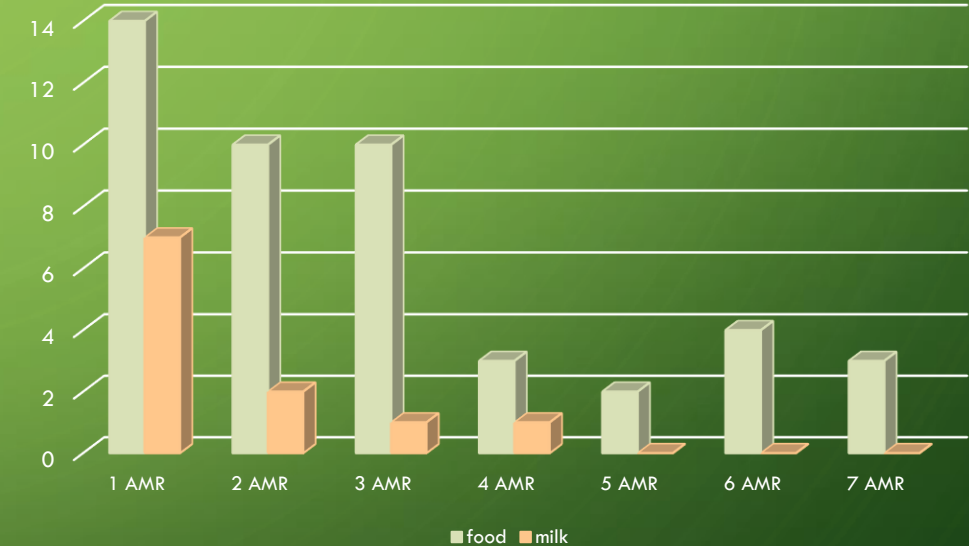
2018-2019: 6 milk/ 46 food strains



*Staphylococcus aureus* resistance for 2018 and 2019



MDR in *S. aureus* for 2018 and 2019



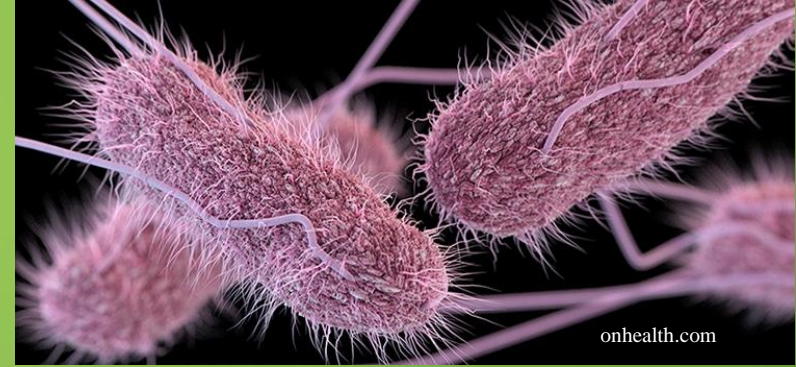
regardless of status (MRSA or not), MDR more pronounced in food strains (*S. aureus* of human origin) than in milk

# DISCUSSION: *CAMPYLOBACTER*



- ❖ Ab treatment usually not required, but effective treatment → shorter illness
- ❖ AMR levels (70%, too high) for ciprofloxacin comparable, although tendencies differ  
→ better don't use anymore in first line (human and vet medicine)
- ❖ Fluoroquinolone resistance persist even after ab pressure removed
- ❖ Macrolide resistance ↓ if antibiotic pressure ↓
- ❖ Reservoirs: ~61% poultry, 33% ruminants, 5% environmental in Luxembourg  
(EFSA → EU: 80% poultry)

# DISCUSSION: *SALMONELLA*



- ❖ Food samples = screening
- ❖ Animal samples: screening and clinical cases
- ❖ Human samples: clinical cases
- ❖ no epidemic diffusion of highly resistant *S. Infantis* or *S. Kentucky* in L
- ❖ *S. Kentucky* resistance to ciprofloxacin and cefotaxime of concern (invasive salmonellosis in humans)
- ❖ Colistin resistance of serious concern
- ❖ Lower levels of AMR in vet medicine (except for Ampicillin)
- ❖ no correlations detectable between human AMR strains and non-human

# DISCUSSION: *E. COLI*



Hospital.vallhebron.com

E.Coli:

|  | ESBL                        | AmpC                    | ESBL + Ampc                 | CarbAp     |
|--|-----------------------------|-------------------------|-----------------------------|------------|
| <u>Cefotaxime</u>                              | R (>1)                      | R (>1)                  | R (>1)                      |            |
| <u>Ceftazidime</u>                             | OU                          | OU                      | OU                          |            |
| <u>Cefoxitine</u>                              | R (>1)                      | R (>1)                  | R (>1)                      |            |
| <u>Meropenem</u>                               | S(<=8)                      | R (>8)                  | R (>8)                      |            |
| <u>Cefotaxime/</u><br><u>Ac. Clavulanique</u>  | S(<=0,12)                   | S(<=0,12)               | S(<=0,12)                   | R (>0,125) |
| <u>Ceftazidime/</u><br><u>Ac. Clavulanique</u> | S (Pos)<br>et/ou<br>S (Pos) | R (Neg)<br>et<br>R(Nég) | S (Pos)<br>et/ou<br>S (Pos) |            |

| Human strains  | Fluoroquinolone > cephalosporin > aminopenicillins   |
|----------------|--|
| Food strains   | aminopenicillins > sulfonamides > tetracyclines > phenicols  |
| Animal strains | Tetracyclines > penicillins > phenicols ( <i>E.coli</i> )<br>β-lactams > sulfonamides > tetracyclines > fluoroquinolones ( <i>E.coli</i> ESBL) |

- ❖ food and animal strains related
- ❖ human and non human strains not related
- ❖ no significant difference between “commensal” *E. coli* and ESBL *E. coli*.



TEM, SHV, CTX, OXA...

- Mainly  $\beta$ -lactamase genes found, some times associations  
→ ampicillin resistance at a rate of 95 to 100%
- cephalosporins 3<sup>rd</sup> generation always affected → difficult treatment
- TEM gene correlated with much higher rates of resistance to amoxicillin-clavulanic acid and piperacillin-tazobactam than with other genes
- Discrepancy between phenotype and genotype



# CONCLUSIONS = LESSONS LEARNT



- AMR = very complex topic ☹
- don't define too quickly couples that won't work
- Interpretation rules change over time → interpretation difficult
- Different or lack of interpretation by matrix and/or default (vet)
- Difficult to compare food-animal-human data
- Human AMR mainly by human source
- Limit ciprofloxacin use
- MDR increasing
- data base !!!
- be careful with low amount of data
- first step taken to develop a One Health approach in Luxembourg



# THANK YOU FOR YOUR ATTENTION

Many thanks to our teams

Mitsou Adam  
Dominique Claude  
Anne Diederich  
Esther Gasperini  
Marilyne Rotondella



Jean-Christophe Even  
Brigitte Martin  
Marie Meo  
Juliette Mirouf  
Alexandre Mzabi  
Anne-Marie Walisch

Serge Losch



Monique Perrin

